

Carbohydrate Polymers 41 (2000) 101-106

Carbohydrate Polymers

www.elsevier.com/locate/carbpol

Phase transition of pectin with sorbed water

M. Iijima^{a,*}, K. Nakamura^a, T. Hatakeyama^a, H. Hatakeyama^b

^aOtsuma Women's University, Sanban-cho, Chiyoda-ku, Tokyo 102-8357, Japan ^bFukui University of Technology, Gakuen, Fukui 910-8505, Japan

Received 13 January 1999; accepted 24 February 1999

Abstract

The phase-transition behaviour of pectin having various degrees of methyl esterification (DE), in the presence of sorbed water, was investigated by differential scanning calorimetry (DSC). In the first run DSC curves of pectin in the dry state, a melting endothermic peak $(T_{\rm m})$ was observed at about 152°C. In the second run DSC curves, glass transition $(T_{\rm g})$ was observed at about 35°C. The above facts indicate that the original pectin formed a regular molecular arrangement. However, amorphous glass is formed when the crystalline pectin is melted once. In pectin—water systems, the $T_{\rm m}$ of pectin decreases with increasing water content $(W_{\rm c}={\rm mass}\ {\rm of}\ {\rm sorbed}\ {\rm water/mass}\ {\rm of}\ {\rm dry}\ {\rm pectin}$ (g/g)). When $W_{\rm c}$ is greater than 0.4 g/g, melting of free water and the glass transition of the pectin—water system were observed. The $T_{\rm g}$ of the pectin—water system decreased with increasing $W_{\rm c}$ in the bound water region. After reaching a minimum value, $T_{\rm g}$ slightly increased and approached a constant value. Water content at the minimum $T_{\rm g}(W_{\rm c_{min}})$ decreased with increasing DE. This suggests that the number of water molecules required to break the hydrogen bonds between –OH and –COOH groups decreases with increasing DE. © 2000 Elsevier Science Ltd. All rights reserved.

Keywords: Pectin; Sorbed water; Phase transition

1. Introduction

Pectin is a complex polysaccharide found in many plant tissues, such as fruits and vegetables (Van Buren, 1991). Pectin consists of 1,4-linked α-D-galacturonic acid having a part of the carboxyl group esterified by a methyl group. The chemical structure of pectin is shown in Fig. 1. The gelation mechanism of pectin varies according to the degree of methyl esterification (DE) (Pedersen, 1980a). When DE is greater than 42.9%, gel is formed in the presence of acids and saccharides. When DE is lesser than 42.9%, gel is formed in the presence of divalent cations. It is known that these two groups of pectins gelatinize by different gelation mechanisms (Thakur, Singh & Handa, 1997). The junction zones of high-methoxyl pectins are stabilized by a combination of hydrogen bonds and hydrophobic interactions between molecules. In contrast, gelation in low-methoxyl pectins occurs by ionic linkages via calcium bridges between two carboxyl groups belonging to two different chains in close contact.

Much research (Al-Ruqaie, Kasapis, Richardson & Mitchell, 1997; Antonov, Lashko & Glotova, 1996; Clark

& Farrer, 1996; Coffin & Fishman, 1994; Garnier, Axelos & Thibault, 1993; Grosso & Rao, 1998; Hoagland & Parris, 1996; Kawabata, 1985; Kawabata & Sawayama, 1975; 1977; Kawabata, Sawayama & Kamata, 1979; Kawabata, Sawayama & Nayoga, 1977; Rao & Cooley, 1994; Willians, Keenan & Halstead, 1998; Yao, Liu, Xiang, Lu & Tu, 1996) on pectins has been carried out, particularly on constituent sugars Kawabata and Sawayama, 1975, solution properties Kawabata and Sawayama, 1977 and mechanism of gelation (Kawabata et al., 1979; Thakur et al., 1997), in the field of food science (Kawabata, 1985), since pectins are used in many foods. Although a large number of studies on pectins have been reported, the physical properties has received little attention (Coffin & Fishman, 1994; Hoagland & Parris, 1996).

Pectins are water-soluble polymers and on this account, it is important to establish the phase-diagram of pectin related to the water content. In our previous studies, we reported that the glass-transition temperature $(T_{\rm g})$ is markedly affected in the presence of bound-water, namely non-freezing water $(W_{\rm nf})$, which is strongly restrained by the hydroxyl group and the ionic group of polysaccharides. In many water—polysaccharide—electrolyte systems, such as water—carboxymethylcellulose, water—cellulose sulfate and water—xanthan gum, $T_{\rm g}$ remarkably decreases by the absorption of non-freezing water. When free water was

^{*} Corresponding author. Tel.: + 81-3-5275-6023; fax: + 81-3-5275-6932.

E-mail address: iijima@otsuma.ac.jp (M. Iijima)

Fig. 1. Chemical structure of pectin. Segment of high-methoxyl pectin, degree of methyl esterification (DE) of 75%.

observed in the system, $T_{\rm g}$ increases, because the molecular motion of polysaccharides associated with bound-water is restricted by ice grown in the intermolecular spaces (Hatakeyama, Nakamura & Hatakeyama, 1996; Nakamura, Hatkeyama & Hatakeyama, 1996).

In this study, the phase-transition behavior of pectins having various DEs, in the presence of non-freezing water and free-water, is investigated by differential scanning calorimetry (DSC).

2. Experimental

2.1. Sample preparation

Pectin in powder-form was obtained from the Taiyo Kagaku Co. DE and viscosity (η) are listed in Table 1. The samples with low DE (P1, P2 and P3) were prepared by de-esterification of the high-methoxyl pectin (P4) (Pedersen, 1980b). It may be noted that high-methoxyl pectin is an intermediate product in the low-methoxyl pectin process. Two methods for de-esterification of acid-demethylated and ammonia-demethylated low-methoxyl pectins are followed in industrial processes. In this experiment, high-methoxyl (P3, P4) and low-methoxyl (P1, P2) pectins were prepared by acid-demethylation and ammonia-demethylation, respectively.

2.2. Measurements

A Seiko Instruments Inc. differential scanning calorimeter EXSTAR 6000, equipped with a cooling apparatus, was used for all the thermal analysis experiments. Temperature and enthalpy calibrations were carried out using indium. When the dry sample was measured, the sample mass was about 10 mg and an aluminium open-pan was used. Dry nitrogen was used as a purge gas and the flow rate was 30 ml/min. The scanning rate was 10°C/min. The

Table 1 Characteristics of pectin samples

Sample		DE (%)	$\eta (m Pa s)^a$
Pectic acid	(P0)	0	
Pectinic acid	(P1)	18-30	20.6
	(P2)	26-36	21.9
	(P3)	64-68	35.9
	(P4)	70–74	166.0

^a $\eta = 2\%$ solution, 25°C.

samples were dried at 120°C for 10 min in a DSC sample-holder. It was confirmed that no endothermic deviation in the sample baseline due to the vaporization of water was observed after heat treatment. The sample was heated to 180°C (first-run measurement), and then rapidly cooled to -150°C . Then the second-run was carried out from -150 to 180°C . Liquid nitrogen was used a coolant.

When the wet sample was measured, the sample weight was 3–5 mg and an aluminium sealed-type sample-pan was used. The sample-pan was hermetically sealed and the total weight of the pectin and water recorded. A Sartorius ultramicro-balance ($\pm 0.1 \times 10^{-6}$ g) was used for sample weight measurements. The accurate water contents (W_c) of the samples were calculated according to the following equation:

$$W_{\rm c} = {\rm mass} \ {\rm of \ sorbed \ water/mass} \ {\rm of \ dry \ pectin} \ ({\rm g/g}).$$
 (1)

The sample pan was pierced with a pin after DSC measurements, in order to remove water from the measured sample. The pan was then dried at 120°C for 90 min. The dried sample was then quickly re-weighed and the intrinsic water content determined.

The melting temperatures ($T_{\rm m}$) of pectin and water were defined as DSC endothermic peaks. $T_{\rm g}$ was defined as the intersection of the extrapolations of the baseline and slope, and the heat capacity difference ($\Delta C_{\rm p}$) was defined between the glassy and the rubbery states at $T_{\rm g}$.

3. Results and discussion

Fig. 2 shows the representative DSC curves of pectin (P4) in the dry state. A melting endothermic peak $(T_{\rm m})$ was observed at 154°C in the first-run. When the sample was cooled from 180 to 0°C and heated from 0 to 180°C (second-run), $T_{\rm g}$ was observed at 37°C. The $T_{\rm g}$ value was constant even if the cooling rate was changed. The crystalline structure of pectin has been investigated by the X-ray diffraction method (Oakenfull, 1991). As shown in Fig. 2, it is clear that the original pectin is a crystalline polymer, but once the crystalline sample is melted, amorphous glass is formed. The amorphous structure of pectin was stable, i.e. the sample was not crystallized by slow cooling or by annealing. It is known that amorphous polysaccharides are crystallized in the presence of water (Hatakeyama, Hatakeyama & Nakamura, 1983). In order to examine the possibility of water-induced crystallization, amorphous pectin

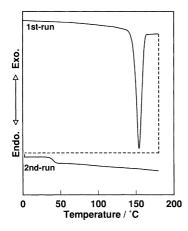


Fig. 2. DSC curves of pectin (P4) powder in the dry state.

was immersed in water at 20°C for 1 month. However, crystallization was not observed.

Fig. 3 shows the relationships between $T_{\rm m}$, melting enthalpy ($\Delta H_{\rm m}$) and DE of pectins in the dry state. $T_{\rm m}$ and $\Delta H_{\rm m}$ increased with increasing DE, indicating that the crystallinity increases with increasing DE. As stated in Section 2, the low-methoxylated pectins were prepared by de-esterification of high-methoxylated pectin. It is thought that the crystalline region of a pectin is destroyed during the chemical reaction.

Fig. 4 shows the relationships between $T_{\rm g}$, $\Delta C_{\rm p}$, estimated from the second-run, and DE of pectins in the dry state. $T_{\rm g}$ maintains a constant value regardless of DE. It is known that bulky side-chains increase intermolecular distance, and main-chain molecular motion is readily enhanced (Nakamura, Hatakeyama & Hatakeyama, 1981). For the samples used in this study, DE values vary, and at the same time, the molecular mass of the pectin decreases with decreasing DE, as shown in Table 1. The high molecular weight introduces chain entanglement that disturbs free molecular rotation. It is thought that $T_{\rm g}$ maintains an almost constant value due to the variate of molecular mass and DE, i.e. molecular mass and bulky side-chain. The slight increase of $\Delta C_{\rm p}$ suggests

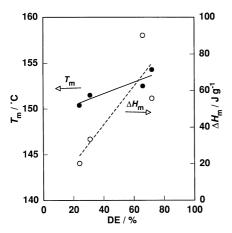


Fig. 3. Relationships between melting temperature (T_m) , melting enthalpy (ΔH_m) and degree of methyl esterification (DE) of pectins in the dry state.

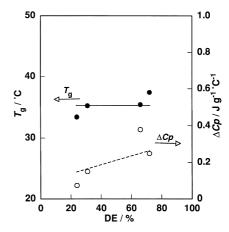


Fig. 4. Relationships between glass transition temperature (T_g) , ΔC_p and degree of methyl esterification (DE) of pectins in the dry state.

that the effect of bulky side-chains is more prominent than the effect of molecular mass.

Fig. 5 shows representative DSC curves of pectin (P4)—water systems having various water contents. Fig. 5(a) shows the heating curves of the first-run and Fig. 5(b) shows those of second-run. Similar DSC curves were obtained in the cases of the other samples (P1, P2 and P3). In the first-run DSC curves, the $T_{\rm m}$ of pectin decreased with increasing $W_{\rm c}$. Glass transition and cold crystallization are clearly observed at $W_{\rm c}=0.437$. At the same time, a small melting peak of ice is observed at a temperature lower than 0°C. Melting enthalpy ($\Delta H_{\rm m}$) of water in the system increases with increasing $W_{\rm c}$. In the second run DSC curves, when $W_{\rm c}$ is small, $T_{\rm g}$ is observed at -50 to 50°C. $T_{\rm g}$ decreased with increasing $W_{\rm c}$. When $W_{\rm c}$ exceeds about 0.5 g/g, the $T_{\rm m}$ of water was observed and $T_{\rm g}$ was constant at about -90°C.

Fig. 6 shows representative phase diagrams of the pectin (P4)—water systems. Similar diagrams were obtained in the other samples. Fig. 6(a) shows the results of the first-run and Part (b), those of the second-run. In the first-run, $T_{\rm m}$ of pectin decreased with increasing $W_{\rm c}$. When $W_{\rm c}$ is greater than 0.4 g/g, $T_{\rm m}$ of sorbed water was observed. When $T_{\rm m}$ of pectin disappeared, $T_{\rm g}$ of pectin was clearly observed. It was found that pectin crystals melt in the presence of more than 0.45 g/g of water.

In the second-run, $T_{\rm g}$ of the pectin decreased remarkably with increasing $W_{\rm c}$ (ranging from 0 to 0.4 g/g). After reaching a minimum value (for example, $-92^{\circ}{\rm C}$ for P4), $T_{\rm g}$ slightly increased and approached a constant value. The minimum value of $T_{\rm g}$ corresponded to the $W_{\rm c}$ where a melting peak of free-water appeared. $\Delta C_{\rm p}$ remarkably increased with increasing $W_{\rm c}$ (ranging from 0 to ca. 1.0 g/g), and $\Delta C_{\rm p}$ decreased when $W_{\rm c}$ increased beyond 1.0 g/g, though the $\Delta C_{\rm p}$ data are not shown in Fig. 6.

Fig. 7(a) shows the variation of $T_{\rm g}$ of pectin-water systems having various DEs and $W_{\rm c}$ s. The variation of $T_{\rm g}$ was magnified in a $W_{\rm c}$ range from 0.4 to 1.0 g/g as shown in Fig. 7(b). $T_{\rm g}$ remarkably decreases in the low $W_{\rm c}$ range, since

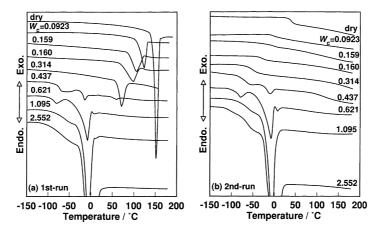


Fig. 5. DSC curves of pectin-water systems containing various water contents (P4): (a) first-run; (b) second-run.

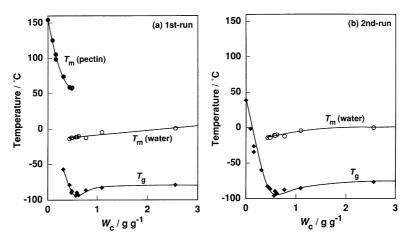


Fig. 6. Phase diagram of the pectin-water system (P4): (a) first-run; (b) second-run.

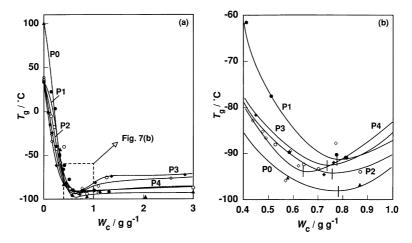


Fig. 7. Relationship between glass transition temperature (T_g) of pectin having various DEs and water content (W_c): (a) W_c ranging from 0 to 3.0 (g/g); (b) W_c ranging from 0.4 to 1.0 (g/g).

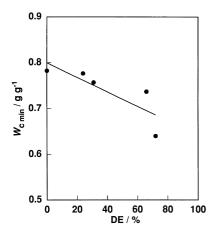


Fig. 8. Relationship between water content at the minimum $T_{\rm g}$ value ($W_{\rm c_{min}}$) and degree of esterification (DE).

intermolecular hydrogen bonding is broken by water molecules. $T_{\rm g}$ values in the initial stage were compared with calculated values derived from Couchman's formula (Couchman, 1978). In the calculation, $T_{\rm g}$ and $\Delta C_{\rm p}$ of dry pectin and those of amorphous ice were used (Sugisaki, Suga & Seki, 1968). The calculated values did not agree well with obtained values. This suggests that bound-water does not exist homogeneously. As shown in Fig. 7(b), $T_{\rm g}$ of pectins shows a minimum value ($T_{\rm g \ min}$) at -91 to $-97^{\circ}{\rm C}$ in the $W_{\rm c}$ ranging from 0.6 to 0.8 g/g. $T_{\rm g \ min}$ shifted to the low $W_{\rm c}$ side with increasing DE.

Fig. 8 shows the relationship between water content at the minimum $T_{\rm g}$ value ($W_{\rm c \ min}$) and DE. $W_{\rm c \ min}$ decreased with increasing DE. When the hydrophilic group (-COCH₃), the extent of intermolecular hydrogen bonding decreases. This causes the number of water molecules necessary to cut the hydrogen bonds between the -OH and -COOH groups to decrease.

4. Conclusion

From the above, the following are concluded: (1) phase diagrams of pectins with various DEs were established in a $W_{\rm c}$ range from 0 to 4.0 g/g; (2) glass-transition temperature of pectin—water systems markedly decreased in a $W_{\rm c}$ range from 0 to 0.4 g/g suggesting that water molecules break intermolecular hydrogen bonding; (3) when ice is formed in the system, $T_{\rm g}$ is constant or slightly increases due to the restriction of free molecular motion by the presence of ice; and (4) the $W_{\rm c}$ showing the lowest $T_{\rm g}$ decreases with increasing DE corresponding to the decrease of the hydroxyl group in the system.

References

Al-Ruqaie, I. M., Kasapis, S., Richardson, R. K., & Mitchell, G. (1997). The glass transition zone in high solids pectin and gellan preparations. *Polymer*, 38, 5685–5694.

- Antonov, Y. A., Lashko, N. P., Glotova, Y. K., Malovikova, A., & Markovich, O. (1996). Effect of the structural features of pectins and alginates on their thermodynamic compatibility with gelatin in aqueous media. Food Hydrocolloids, 10, 1–9.
- Clark, A. H., & Farrer, D. B. (1996). Shear modulus—concentration relationships for low DE pectin—calcium gels in the temperature range 20-85(C. Food Hydrocolloids, 10, 31–39.
- Coffin, D. R., & Fishman, M. L. (1994). Physical and mechanical properties of highly plasticized pectin/starch films. *Journal of Applied Polymer Science*, 54, 1311–1320.
- Couchman, P. R. (1978). Compositional variation of glass-transition temperature. 2. Application of the thermodynamic theory to compatible polymer blends. *Macromolecules*, 11, 1156–1161.
- Garnier, C., Axelos, M. A. V., & Thibault, J. -F. (1993). Phase diagrams of pectin–calcium systems, influence of pH, ionic strength, and temperature on the gelation of pectins with different degree of methoxylation. *Carbohydrate Research*, 240, 219–232.
- Grosso, C. R. F., & Rao, M. A. (1998). Dynamic rheology of structure development in low-methoxyl pectin + Ca²⁺ + sugar gels. Food Hydrocolloids, 12, 357–363.
- Hatakeyama, H., Hatakeyama, T., & Nakamura, K. (1983). Relationship between hydrogen bonding and water in cellulose. *Journal of Applied Polymer Science Symposia*, 37, 979.
- Hatakeyama, T., Nakamura, K., & Hatakeyama, H. (1996). Glass transition of polysaccharide electrolyte-water systems. *Kobunshi Robunshu*, 53, 795–802.
- Hoagland, P. D., & Parris, N. (1996). Chitosan/pectin laminated films. Journal of the Agricultural Food Chemistry, 44, 1915–1919.
- Kawabata, A. (1985). Studies on pectic substances from the view point of cookery science—chemical and physical properties of pectic substances from fruits. *Journal of Home Economy in Japan*, 36, 561– 576.
- Kawabata, A., & Sawayama, S. (1975). Constituent sugars of pectic substances from fruits. *Journal of Japanese Society of Food and Nutri*tion, 28, 395–402.
- Kawabata, A., & Sawayama, S. (1977). Molecular weights and molecular size of pectins. *Journal of the Agricultural Chemical Society of Japan*, 51, 15–21.
- Kawabata, A., Sawayama, S., & Nagoya, T. (1977). Viscosity behavior of solutions. *Journal of Japanese Society of Food and Nutrition*, 30, 149– 154.
- Kawabata, A., Sawayama, S., & Kamata, T. (1979). On the mechanism of association of low-methoxyl pectin due to calcium ions. *Journal of the Agricultural Chemical Society of Japan*, 53, 61–67.
- Nakamura, K., Hatakeyama, T., & Hatakeyama, H. (1981). Differential scanning calorimetric studies on the glass transition temperature of polyhydroxystyrene derivatives containing sorbed water. *Polymer*, 22, 473–476.
- Nakamura, K., Hatakeyama, T., & Hatakeyama, H. (1996). Heat capacities of carboxymethylcellulose–nonfreezing water systems at around glass transition temperature. *Kobunshi Robunshu*, 53, 860–865.
- Oakenfull, D. G. (1991). The chemistry of high-methoxyl pectins. In R. H. Walter (Ed.), *The chemistry and technology of pectin*, (pp. 96–97). New York: Academic Press.
- Pedersen, J. K. (1980). Pectins. In R. L. Davidson (Ed.), Handbook of water-soluble gums and resins, (pp. 15/1–15/5). New York: McGraw Hill.
- Pedersen, J. K. (1980). Pectins. In R. L. Davidson (Ed.), Handbook of water-soluble gums and resins, (pp. 15/6). New York: McGraw-Hill.
- Rao, M. A., & Cooley, H. J. (1994). Influence of glucose and fructose on high-methoxyl pectin gel strength and structure development. *Journal* of Food Quality, 17, 21–31.
- Sugisaki, M., Suga, H., & Seki, S. (1968). Calorimetric study of the glassy state. IV. Capacities of glassy water and cubic ice. *Bulletin of Chemical Society of Japan*, 41, 2591–2599.
- Thakur, B. R., Singh, R. K., & Handa, A. K. (1997). Chemistry and uses of

- pectin—a review. Critical Reviews in Food Science and Nutrition, 37, 47–73.
- Van Buren, J. P. (1991). Function of pectin in plant tissue structure and firmness. In R. H. Walter (Ed.), *The chemistry and technology of pectin* (pp. 1–2). New York: Academic Press pp. 1–2.
- Willians, M. A. K., Keenan, R. D., & Halstead, T. K. (1998). ²H NMR
- lineshape study of $-CH_3$ group dynamics in pectin gels. Food Hydrocolloids, 12, 89–94.
- Yao, K. D., Liu, J., Xiang, G., Lu, X. D., & Tu, H. L. (1996). Swelling behavior of pectin/chitosan complex films. *Journal of Applied Polymer Science*, 60, 279–283.